

LIPOLYTIC CAPACITY OF *Pseudomonas* spp. ISOLATED FROM REFRIGERATED RAW MILK

CAPACIDADE LIPOLÍTICA DE *Pseudomonas* spp. ISOLADAS DE LEITE CRU REFRIGERADO

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ABSTRACT: The psychrotrophic bacteria growth in refrigerated milk is responsible for the production of heat resistant enzymes, including those which have lipolytic capacity. Lipases can cause rancidity and off-flavor in pasteurized and UHT milk, milk powder and cheese. *Pseudomonas* spp. is the major genera responsible by lipases synthesis, mainly *Pseudomonas fluorescens*. The lipolytic capacity of *Pseudomonas* spp. (1182 strains) and *P. fluorescens* (158 strains) isolated from cooling tank and bulk milk transportation were tested. *Pseudomonas* spp. strains were isolated by Pseudomonas Agar Base, adding the supplement cephalotin, fusidic acid, cetrimide- CFC (30° C/48 h) and Pseudomonas Cetrimide Agar with 10% of glicerol (21° C/ 48 h) to determine *P. fluorescens*. The strains lipolytic capacity were tested with Tributyrin Agar (21 °C/72 h). Lipolytic indexes of *Pseudomonas* spp. were 51.4% and 67.2% in cooling tank and bulk milk transportation ($p < 0.05$), respectively. *P. fluorescens* lipolytic capacity indexes were 26.4% from strains isolated from cooling tank and 41% ($p < 0.05$) from strains isolated from bulk milk transportation. High lipolytic indexes had been found in the strains isolated from refrigerated raw milk sent to manufacturing. Refrigeration temperature and storage time may contribute to the difference observed between two points of collection evaluated in this study.

KEYWORDS: Psychrotrophic. Enzymes. Quality.

INTRODUCTION

High counts of psychrotrophic bacteria in raw milk are directly related to poor hygienic conditions during production and milking, and to the time and temperature of milk storage (ARCURI et al., 2008). Low psychrotrophic bacteria counts are essential to maintain milk quality, once the metabolic activity of these microorganisms modify milk biochemical constituents (ARCURI et al., 2008), such as lipid, protein and carbohydrate degradation (COUSIN, 1982), limiting the shelf life of fluid milk and its products (SANTOS et al. 2009; ARCURI et al., 2008).

Lipolysis involves the enzymatic hydrolysis of milk lipids, producing free fatty acids and glycerol. Lipases can be of endogen origin, being the lipoprotein lipase the most important, or exogenous, produced by microbial activity. Bacterial lipases are not inactivated by heating, and cause rancidity and off-flavor in pasteurized and UHT milk, milk powder and cheese (JONGHE et al., 2011; COUSIN, 1982). *Pseudomonas* spp. is the major genera responsible by lipases synthesis, mainly *Pseudomonas fluorescens* (MU et al., 2008; DEETH; FITZ-GERALD, 2006; SHAH, 1994).

Thus, this study aimed to evaluate the lipolytic capacity of *Pseudomonas* spp. strains and

P. fluorescens isolated from refrigerated raw milk from cooling tanks of rural properties and from bulk milk transportation responsible for the raw milk transportation to the dairy industry.

MATERIAL AND METHODS

Raw milk samples were collected between June 2013 and February 2014. A total of 37 samples were collected from cooling tanks (CT) from eight rural properties from Paraná State, Brazil, 48 h after the milking. Samples from bulk milk transportation (BMT) of the same rural properties were also collected, in a total of nine samples. The milk temperature indicated in equipment was recorded.

Pseudomonas spp. strains were isolated by Pseudomonas Agar Base, adding the supplement cephalotin, fusidic acid, cetrimide (CFC) (Himedia, Mumbai, Índia), at 30° C/48 h (FAGUNDES et al., 2006). To determine *P. fluorescens*, Pseudomonas Cetrimide Agar (Himedia, Mumbai, India) was incubated at 21° C/ 48 h (SANTOS et al., 2009; KING, 1954). To verify the *Pseudomonas* spp. and *P. fluorescens* strains isolated from milk samples lipolytic capacity, plates with maximum of 100 colony formatting units were selected, and considered positive the colonies with transparent halo. Thus, 1182 *Pseudomonas* spp. strains and 158

P. fluorescens strains were tested. Positive colonies were plated on Tributyrin Agar (Himedia, Mumbai, India) and incubated at 21 °C/72 h (FRANK et al, 1992).

Differences in lipolytic capacity of strains isolated from CT and BMT were tested by Chi-square test ($p < 0.05$). The null hypothesis tested was that the lipolytic strains isolated from CT frequency was not different from that of BMT. Thus, the alternative hypothesis tested was that the lipolytic strains isolated from CT frequency was the same from that of BMT.

RESULTS AND DISCUSSION

From *Pseudomonas* spp. strains isolated from CT milk, 51.4% presented lipolytic capacity. Among *Pseudomonas* spp. strains isolated from BMT, a higher number of 67.2% presented lipolytic capacity (Table 1). Lipases are mainly produced at 21 °C, but they also occur at refrigeration temperature, during the log phase final of cellular

growing (MAHIEU, 1991). This fact can explain the difference ($p < 0.05$) observed in this study in the lipolytic strains frequency from CT and BMT (Table 1). The result of Chi-square test is to reject the null hypothesis, that is, the lipolytic strains frequency isolated from CT was different from that of BMT. At low temperature, *Pseudomonas* spp. had a short lag phase and a long stationary phase, and can survive for extended periods in milk residues (SORHAUNG; STEPANIAK 1997). Kumaresan et al. (2007) observed during 14 days a difference in lipolytic activity at 4 °C and 7 °C, with lower lipolytic capacity in temperatures under 4 °C. According to these authors, the enzymatic activity also depends on the raw milk refrigeration time, tested at 0, 3, 5, 7 and 14 days of storage. In the present study, a similar result was obtained. Difference ($p < 0.05$) between the number of lipolytic strains in milk stored during 48 h (CT), with mean temperature of 3.8 °C, and the raw milk delivered to the dairy industry (BMT), with mean temperature of 6.6 °C was observed (Table 1).

Table 1. Lipolytic capacity of *Pseudomonas* spp. and *P. fluorescens* strains isolated from raw refrigerated milk from cooling tanks (CT) of rural properties and from bulk milk transportation (BMT) responsible for the raw milk transportation to the dairy industry.

Sample	T°C	<i>Pseudomonas</i> spp strains			<i>P. fluorescens</i> strains		
		TS	LS	LS (%)	TS	LS	LS (%)
CT	3.8	1002	515	51.4 ^b	129	34	26.4 ^b
BMT	6.6	180	121	67.2 ^a	29	12	41.0 ^a
Total	---	1182	636	53.8	158	46	29.1

Values with different lower cases in the same column differ significantly by the Chi-Square test ($p < 0.05$); CT = raw refrigerated milk from cooling tanks; BMT = raw refrigerated milk from bulk milk transportation. TS= Tested strains; LS= Lipolytic strains.

The use of refrigeration without good practices during the raw production allows the psychrotrophic microorganisms growing that produces heat resistant lipases (IZIDORO et al., 2013). At low temperatures (4 °C), the active transportation and the inner nutrients diffusion to bacterial cells are reduced. To compensate this mechanism, enzymatic production is increased, including lipases production (BUCK et al., 1986).

When the *P. fluorescens* strains lipolytic capacity in the raw refrigerated milk from cooling tanks (3.8 °C) was compared to that from bulk milk transportation (6.6 °C), we observed indexes of 26.4% and 41%, respectively (Table 1). Different results were observed by Arcuri et al. (2008), where all *P. fluorescens* strains isolated at 4 °C, 7 °C, 10 °C and 21 °C had lipolytic capacity. The enzymatic production of lipases, proteases and phospholipases is related with the temperature, microorganism

growing phase, oxygen availability and medium composition. Its activity depends on temperature, pH and substrate concentration (NUÑEZ; NUÑEZ, 1983).

Milk thermal treatment such as pasteurization and ultra high temperature (UHT) reduces the lipolytic activity in 75% and 91.6%, respectively. However, extracellular lipases produced by psychrotrophic bacteria resist to sterilization at 130 °C/15 seconds (SHAH, 1994). The majority of bacterial lipases had specificity to sn-1 and sn-3 positions of the triacylglycerol, and can hydrolyse monoacylglycerols and diacylglycerols at higher speed when compared to triacylglycerols (ARCURI et al., 2006).

Besides rancidity and bitter taste, other problems and defects are caused due the *Pseudomonas* spp. enzymatic activity such as UHT milk gelation, fruit flavor (JONGHE et al., 2011;

MU et al., 2009), thermal stability, ethanol stability, false positive result to the search of milk fraud by wheat addition through the sialic acid determination and reduction in the cheese yielding (ARCURI et al., 2008).

CONCLUSION

In the refrigerated raw milk studied, high lipolytic capacity indexes of *Pseudomonas* spp. and *P. fluorescens* strains were observed, with higher values in milk from bulk milk transportation delivered to the dairy industry. Refrigeration temperature and storage time may contribute to the difference observed in the two collection points.

RESUMO: A multiplicação de bactérias psicotróficas em leite refrigerado pode promover a síntese de enzimas resistentes ao calor, entre elas aquelas que tem capacidade lipolítica. As lipases podem promover rancidez, e defeitos de sabor em leite pasteurizado, UHT, leite em pós e queijos. *Pseudomonas* spp. é o principal gênero responsável pela síntese desta enzimas, principalmente a espécie *Pseudomonas fluorescens*. Avaliou-se a capacidade lipolítica de 1182 cepas de *Pseudomonas* spp. e 158 cepas de *P. fluorescens* isoladas de amostras de leite de tanques de resfriamento (TR) e caminhão tanque (CT) enviado ao beneficiamento. Foi utilizado o Agar base *Pseudomonas* com suplemento cephalotin, fusidic acid, cetrimide - CFC (30° C/48 h) para isolamento das pseudomonas e *Pseudomonas* Cetrimide Agar com 10% de glicerol (21° C/ 48 h) para a espécie *P. fluorescens*. Para avaliação da capacidade lipolítica as cepas foram semeadas em Ágar Tributirina (21 °C/72 h). O índice de lipólise das cepas de *Pseudomonas* spp. provenientes do leite do tanque de resfriamento e do caminhão tanque (p<0,05) foram de 51,4% e 62,7%, respectivamente. Foram também observados índices de capacidade lipolítica de *P. fluorescens* de 26,4% e 41% (p<0,05) para TR e CT, respectivamente. Altos índices de cepas lipolíticas foram encontrados nas cepas isoladas do leite cru refrigerado enviado ao beneficiamento. Temperatura de refrigeração e tempo de estocagem podem ter contribuído para a diferença observada na população lipolítica dos dois pontos de coleta avaliados.

PALAVRAS-CHAVE: Psicotróficos. Enzimas. Qualidade.

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